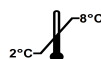


## Instructions for use

# Glycine ELISA

**REF**

**BA E-2100**



**RUO**

For research  
use only –  
Not for use  
in diagnostic  
procedures

## Glycine Urine ELISA

### 1. Intended use and principle of the test

Enzyme Immunoassay for the quantitative determination of Glycine in urine.

After derivatization Glycine is quantitatively determined by ELISA. The competitive ELISA uses the microtiter plate format. The antigen is bound to the solid phase of the microtiter plate. The derivatized standards, controls and samples and the solid phase bound analyte compete for a fixed number of antiserum binding sites. When the system is in equilibrium, free antigen and free antigen-antiserum complexes are removed by washing. The antibody bound to the solid phase is detected by an anti-rabbit IgG-peroxidase conjugate using TMB as a substrate. The reaction is monitored at 450 nm.

Quantification of unknown samples is achieved by comparing their absorbance with a reference curve prepared with known standards.

### 2. Procedural Cautions, Guidelines and Warnings

- (1) This kit is intended for professional use only. Users should have a thorough understanding of this protocol for the successful use of this kit. Only the test instruction provided with the kit is valid and has to be used to run the assay. Reliable performance will only be attained by strict and careful adherence to the instructions provided.
- (2) The principles of Good Laboratory Practice (GLP) have to be followed.
- (3) In order to reduce exposure to potentially harmful substances, wear lab coats, disposable latex gloves and protective glasses where necessary.
- (4) All kit reagents and specimens should be brought to room temperature and mixed gently but thoroughly before use. Avoid repeated freezing and thawing of reagents and specimens.
- (5) For dilution or reconstitution purposes, use deionized, distilled, or ultra-pure water.
- (6) The microtiter plate contains snap-off strips. Unused wells must be stored at 2 °C to 8 °C in the sealed foil pouch with desiccant and used in the frame provided.
- (7) Duplicate determination of sample is highly recommended to be able to identify potential pipetting errors.
- (8) Once the test has been started, all steps should be completed without interruption. Make sure that the required reagents, materials and devices are prepared ready at the appropriate time.
- (9) Incubation times do influence the results. All wells should be handled in the same order and time intervals.
- (10) To avoid cross-contamination of reagents, use new disposable pipette tips for dispensing each reagent, sample, standard and control.
- (11) A standard curve must be established for each run.
- (12) The controls should be included in each run and fall within established confidence limits. The confidence limits are listed in the QC-Report.
- (13) Do not mix kit components with different lot numbers within a test and do not use reagents beyond expiry date as shown on the kit labels.
- (14) Avoid contact with Stop Solution containing 0.25 M H<sub>2</sub>SO<sub>4</sub>. It may cause skin irritation and burns. In case of contact with eyes or skin, rinse off immediately with water.
- (15) TMB substrate has an irritant effect on skin and mucosa. In case of possible contact, wash eyes with an abundant volume of water and skin with soap and abundant water. Wash contaminated objects before reusing them.
- (16) For information on hazardous substances included in the kit please refer to Safety Data Sheet (SDS). The Safety Data Sheet for this product is made available directly on the website of the manufacturer or upon request.
- (17) Kit reagents must be regarded as hazardous waste and disposed according to national regulations.

### 3. Storage and stability

Store the unopened reagents at 2 - 8 °C until expiration date. Do not use components beyond the expiry date indicated on the kit labels. Once opened the reagents are stable for 1 month when stored at 2 - 8 °C. Once the resealable pouch has been opened, care should be taken to close it tightly with desiccant again.

### 4. Materials

#### 4.1 Content of the kit

<b>BA D-0090</b>	<b>FOILS</b>	<b>Adhesive Foil</b> - Ready to use
Content:	Adhesive Foils in a resealable pouch	
Volume:	1 x 4 foils	

**BA E-0030** **WASH-CONC 50x** **Wash Buffer Concentrate** - Concentrated 50x

Content: Buffer with a non-ionic detergent and physiological pH

Volume: 1 x 20 ml/vial, light purple cap

**BA D-0024** **REAC-PLATE** **Reaction Plate** - Ready to use

Content: 1 x 96 well plate, empty in a resealable pouch

**BA E-0040** **CONJUGATE** **Enzyme Conjugate** - Ready to use

Content: Goat anti-rabbit immunoglobulins conjugated with peroxidase

Volume: 1 x 12 ml/vial, red cap

**BA E-0055** **SUBSTRATE** **Substrate** - Ready to use

Content: Chromogenic substrate containing tetramethylbenzidine, substrate buffer and hydrogen peroxide

Volume: 1 x 12 ml/black vial, black cap

**BA E-0080** **STOP-SOLN** **Stop Solution** - Ready to use

Content: 0.25 M sulfuric acid

Volume: 1 x 12 ml/vial, light grey cap

Hazards identification:



H290 May be corrosive to metals.

H314 Causes severe skin burns and eye damage.

**BA E-2413** **ASSAY-BUFF** **Assay Buffer** - Ready to use

Contents: Buffer with alkaline pH

Volume: 1 x 20 ml/vial, yellow cap

**BA E-2428** **EQUA-REAG** **Equalizing Reagent** - Lyophilized

Contents: Lyophilized protein

Volume: 1 vial, brown cap

**Standards and Controls** - Ready to use

Cat. no.	Component	Colour/Cap	Concentration µg/ml	Concentration µmol/l	Volume/ Vial
<b>BA E-2101</b>	<b>STANDARD A</b>	white	0	0	4 ml
<b>BA E-2102</b>	<b>STANDARD B</b>	light yellow	10	133	4 ml
<b>BA E-2103</b>	<b>STANDARD C</b>	orange	30	399	4 ml
<b>BA E-2104</b>	<b>STANDARD D</b>	dark blue	100	1 330	4 ml
<b>BA E-2105</b>	<b>STANDARD E</b>	light grey	300	3 990	4 ml
<b>BA E-2106</b>	<b>STANDARD F</b>	black	1 000	13 300	4 ml
<b>BA E-2151</b>	<b>CONTROL 1</b>	light green	Refer to QC-Report for expected value and acceptable range!		4 ml
<b>BA E-2152</b>	<b>CONTROL 2</b>	dark red			4 ml

Conversion: Glycine (µg/ml) x 13.3 = Glycine (µmol/l)

Contents: Acidic buffer with non-mercury preservative, spiked with defined quantity of Glycine

**BA E-2131** **GLY** **Glycine Microtiter Strips** - Ready to use

Contents: 1 x 96 well (12x8) antigen precoated microwell plate in a resealable foil pouch with desiccant

**BA E-2110** **ASGLY** **Glycine Antiserum** - Ready to use

Contents: Rabbit anti- glycine antibody, blue coloured

Volume: 1 x 6 ml/vial, blue cap

**BA E-2446** D-REAGENT **D-Reagent - Ready to use**

Contents: Crosslinking agent in dimethylsulfoxide

Volume: 1 x 4 ml/vial, white cap

**BA E-2129** RED-CONC 100x **Reducing Concentrate - Ready to use**

Contents: Reducing agent in sodium hydroxide

Volume: 1 x 1 ml/vial, pink cap

Hazards identification:



H290 May be corrosive to metals.

H301 Toxic if swallowed.

H314 Causes severe skin burns and eye damage.

H360FD May damage fertility. May damage the unborn child.

#### 4.2 Additional materials and equipment required but not provided with the kit

- Calibrated precision pipettes to dispense volumes between 20 µl – 300 µl; 2,5 ml; 12,5 ml
- Polystyrene or polypropylene tubes and suitable rack
- Microtiter plate washing device (manual, semi-automated or automated)
- ELISA reader capable of reading absorbance at 450 nm and if possible 620 - 650 nm
- Microtiter plate shaker (shaking amplitude 3 mm; approx. 600 rpm)
- Absorbent material (paper towel)
- Water (deionized, distilled, or ultra-pure)
- Vortex mixer

#### 5. Sample collection and storage

##### Urine

Spontaneous or 24-hour urine, collected in a bottle containing 10 – 15 ml of 6 M HCl, should be used.

Determine the total volume of urine excreted during a period of 24 h for calculation of the results.

Storage: for longer periods (up to 6 months) at -20 °C.

Repeated freezing and thawing should be avoided.

Avoid exposure to direct sunlight.

#### 6. Test procedure

Allow all reagents to reach room temperature and mix thoroughly by gentle inversion before use. Duplicate determinations are recommended. It is recommended to number the strips of the microwell plate before usage to avoid any mix-up.

The binding of the antibodies and the enzyme conjugates and the activity of the enzyme used are temperature dependent, and the absorption values may vary if a thermostat is not used. The higher the temperature, the higher the absorption values will be. The absorption values also depend on the incubation times. The optimal temperature during the Enzyme Immunoassay is between 20 – 25 °C.

*In case of overflow, read the absorbance of the solution in the wells within 10 minutes, using a microplate reader set to 405 nm.*

#### 6.1 Preparation of reagents

##### Wash Buffer

Dilute the 20 ml Wash Buffer Concentrate with water (deionized, distilled, or ultra-pure) to a final volume of 1000 ml.

Storage: 1 month at 2 - 8 °C

##### Equalizing Reagent

Reconstitute the Equalizing Reagent with **12.5 ml** of **Assay Buffer**.

Reconstituted Equalizing Reagent which is not used immediately has to be stored in aliquots for max 1 month at -20 °C and may be thawed only once.

##### D-Reagent

The D-Reagent has a freezing point of 18.5 °C. To ensure that the D-Reagent is liquid when being used, it must be ensured that the D-Reagent has reached room temperature and forms a homogeneous, crystal-free solution.

### Reducing Solution

Dilute **Reducing Concentrate** 1:100 with **water** (deionized, distilled, or ultra-pure) and mix thoroughly. Use immediately!

Examples for the preparation of Reducing Solution:

<b>Reducing Concentrate</b>	40 µl	50 µl	80 µl	160 µl
<b>Water</b>	3.96 ml	4.95 ml	7.92 ml	15.84 ml

### Glycine Microtiter Strips

In rare cases residues of the blocking and stabilizing reagent can be seen in the wells as small, white dots or lines. These residues do not influence the quality of the product.

## 6.2 Dilution

1. Pipette <b>20 µl</b> of <b>standards, controls</b> and <b>samples</b> into the respective <b>tubes</b> .
2. Add <b>2.5 ml</b> of <b>water</b> (deionized, distilled, or ultra-pure) to all tubes and mix thoroughly (vortex).
▲ Take <b>100 µl</b> for the <b>derivatization</b> .

## 6.3 Derivatization

1. Pipette <b>100 µl</b> of the <b>diluted standards, controls</b> and <b>samples</b> into the appropriate wells of the <b>Reaction Plate</b> .
2. Add <b>50 µl</b> of the <b>Equalizing Reagent</b> to all wells.
4. Add <b>10 µl</b> of the <b>D-Reagent</b> to all wells.
5. Cover plate with <b>Adhesive Foil</b> and shake for <b>2 h</b> at <b>RT</b> (20 - 25 °C) on a <b>shaker</b> (approx. 600 rpm).
6. Pipette <b>150 µl Reducing Solution</b> (refer to 6.1) into all wells. <b><i>The Reducing Solution should be prepared directly prior to use!</i></b>
7. Shake for <b>30 min</b> at <b>RT</b> (20 - 25 °C) on a <b>shaker</b> (approx. 600 rpm).
▲ Take <b>25 µl</b> for the <b>ELISA!</b>

## 6.4 Glycine ELISA

1. Pipette <b>25 µl</b> of the <b>prepared standards, controls and samples</b> into the appropriate wells of the <b>Glycine Microtiter Strips</b> .
2. Pipette <b>50 µl</b> of the <b>Glycine Antiserum</b> into all wells and mix shortly.
3. Cover plate with <b>Adhesive Foil</b> and incubate for <b>15 - 20 h</b> (overnight) at <b>2 - 8 °C</b> .
4. Remove the foil. Discard or aspirate the contents of the wells. Wash the plate <b>3 x</b> by adding <b>300 µl</b> of <b>Wash Buffer, discarding</b> the content and <b>blotting dry each time</b> by tapping the inverted plate on absorbent material.
5. Pipette <b>100 µl</b> of the <b>Enzyme Conjugate</b> into all wells.
6. Incubate for <b>30 min</b> at <b>RT</b> (20 - 25 °C) on a <b>shaker</b> (approx. 600 rpm).
7. Remove the foil. Discard or aspirate the contents of the wells. Wash the plate <b>3 x</b> by adding <b>300 µl</b> of <b>Wash Buffer, discarding</b> the content and <b>blotting dry each time</b> by tapping the inverted plate on absorbent material.
8. Pipette <b>100 µl</b> of the <b>Substrate</b> into all wells and incubate for <b>20-30 min</b> at <b>RT</b> (20 - 25 °C) on a <b>shaker</b> (approx. 600 rpm). <b><i>Avoid exposure to direct sunlight!</i></b>
9. Add <b>100 µl</b> of the <b>Stop Solution</b> to each well and shake the microtiter plate to ensure a homogeneous distribution of the solution.
10. <b>Read</b> the absorbance of the solution in the wells within 10 minutes, using a microplate reader set to <b>450 nm</b> (if available a reference wavelength between 620 nm and 650 nm is recommended).

**7. Calculation of results**

<b>Measuring range</b>	<b>Glycine</b>
	3.3 – 1 000 µg/ml

The standard curve is obtained by plotting the absorbance readings (calculate the mean absorbance) of the standards (linear, y-axis) against the corresponding standard concentrations (logarithmic, x-axis). Use non-linear regression for curve fitting (e.g. spline, 4- parameter, akima).

*This assay is a competitive assay. This means: the OD-values are decreasing with increasing concentrations of the analyte. OD-values found below the standard curve correspond to high concentrations of the analyte in the sample and have to be reported as being positive.*

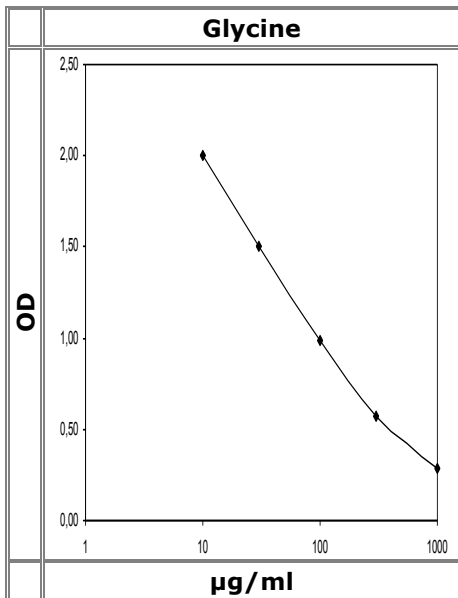
The concentrations of the **samples** and **controls** can be read directly from the standard curve.

**7.1 Quality control**

The confidence limits of the kit controls are indicated on the QC-Report.

**7.2 Typical standard curve**

*Example, do not use for calculation!*



**8. Assay characteristics**

<b>Analytical Sensitivity (Limit of Detection)</b>	<b>Glycine</b>
	3.3 µg/ml

<b>Analytical Specificity (Cross Reactivity)</b>	<b>Substance</b>	<b>Cross Reactivity (%)</b>
	Glycine	100
	D-Serin	3.7
	L-Cystein	1.8
	Beta-Alanin	0.7
	GABA	0.8
	L-Aspartic Acid	<0.1
	L-Glutamat	<0.1
	Taurin	<0.1







<b>Precision</b>					
<b>Intra-Assay</b>			<b>Inter-Assay</b>		
Sample	Range (µg/ml)	CV (%)	Sample	Range (µg/ml)	CV (%)
1 (n = 20)	66.7 ± 4.2	6.2	1 (n = 23)	63.3 ± 7.9	13
2 (n = 20)	94.0 ± 3.5	3.7	2 (n = 23)	91.1 ± 9.2	10
3 (n = 20)	217 ± 11.0	5.1	3 (n = 23)	211 ± 9.4	9.4

<b>Linearity</b>		Range Linearity %	Serial dilution up to	Mean Linearity %
	Urine	94 - 116	1:128	100

<b>Recovery</b>		Mean Creatinine (mg/dl)	Mean Recovery (%)	Range Recovery (%)
	Sample 1	38.8	108	96 - 116
	Sample 2	102	95	93 - 96
	Sample 3	135	108	105 - 114

- **For literature or any other information please contact your local supplier.**
- **The liability of the manufacturer shall be limited to the replacement of defective products. The manufacturer takes no liability for any damages or expenses arising directly or indirectly from the use of this product.**

**Symbols:**

	Storage temperature		Manufacturer		Contains sufficient for <n> tests
	Expiry date	<b>LOT</b>	Batch code		
	Consult instructions for use	<b>CONT</b>	Content		
	Caution	<b>REF</b>	Catalogue number	<b>RUO</b>	For research use only!