

Instructions for use
HistaSure™ ELISA Fast Track



REF

FC E-3600



1. Intended use and principle of the test

The **HistaSure™ ELISA** Fast Track is intended for the rapid semi-quantitative or quantitative determination of histamine in different scombroid fish types such as tuna, mahi mahi, sardines and for the determination of histamine in fishmeal.

The assay kit provides materials for the determination of derivatized histamine in food extracts. The derivatization is part of the preparation of the samples. By use of the acylation reagent, histamine is quantitatively derivatized into N-acylhistamine. The competitive Histamine ELISA kit uses the microtiter plate format. Histamine is bound to the solid phase of the microtiter plate. Acylated histamine and solid phase bound histamine compete for a fixed number of antiserum binding sites. When the system is in equilibrium, free antigen and free antigen-antiserum-peroxidase complexes are removed by washing. The substrate TMB/peroxidase reaction is monitored at 450 nm. The amount of antibody bound to the solid phase histamine is inversely proportional to the histamine concentration of the sample.

2. Introduction

Histamine testing in fresh fish is a possible control strategy that can be used by seafood processors in their HACCP program to address the hazard of scombrototoxin formation. Histamine is a product of decomposition of histidine caused by the growth of certain bacteria in seafood. The amount of the amine that forms is a function of bacterial species, the temperature and time of exposure, and may exceed 1,000 ppm (mg/kg). Fish containing high levels of histamine has been associated with many examples of poisoning commonly referred to as "scombroid poisoning," a major health problem for consumers. Scombrototoxic fish usually contains levels of histamine in excess of 200 ppm but such fish may be randomly dispersed within a lot. For large fish, histamine is found at variable levels even within individual fish. Quality control measures designed to minimize the occurrence of scombrototoxic fish require the determination of histamine levels in the range of approximately 10 to 200 ppm. Good quality fish contain less than 10 ppm histamine, a level of 30 ppm indicates significant deterioration, and 50 ppm is considered to be evidence of definite decomposition. The defect action level (DAL), the level at which regulatory actions are taken for histamine is 50 ppm (P. L. Rogers, W. F. Staruszkiewicz, Journal of Aquatic Food Product Technology, Vol. 9 (2) 2000 p. 5 - 17).

3. Procedural Cautions, Guidelines and Warnings

- This kit is for professional use only. Users should have a thorough understanding of this protocol for the successful use of this kit. Only the test instruction provided with the kit is valid and has to be used to run the assay. Reliable performance will only be attained by strict and careful adherence to the instructions provided.
- In order to reduce exposure to potentially harmful substances, wear lab coats, disposable latex gloves and protective glasses where necessary.
- All kit reagents and specimens should be brought to room temperature (20 - 25 °C) and mixed gently but thoroughly before use.
- When the use of water is specified for dilution or reconstitution, use deionized, distilled, or ultra-pure water.
- The microplate contains snap-off strips. Unused wells must be stored at 2 °C to 8 °C in the sealed foil pouch and used in the frame provided. Wells are for single use only.
- Once the test has been started, all steps should be completed without interruption. Make sure that required reagents, materials and devices are prepared ready at the appropriate time.
- Incubation times do influence the results. All wells should be handled in the same order and time sequences.
- To prevent contamination of reagents, use a new disposable pipette tip for dispensing each reagent, sample, standard and control.
- Do not mix various lot numbers of kit components within a test and do not use reagents beyond expiry date as shown on the kit labels.
- Avoid contact with Stop Solution containing 0.25 M H₂SO₄. It may cause skin irritation and burns. In case of contact with eyes or skin, flush immediately with water.
- Some reagents contain sodium azide (NaN₃) as preservatives. In case of contact with eyes or skin, flush immediately with water. NaN₃ may react with lead and copper plumbing to form explosive metal azides. When disposing reagents, flush with a large volume of water to avoid azide build-up.
- TMB substrate has an irritant effect on skin and mucosa. In case of possible contact, wash eyes with an abundant volume of water and skin with soap and abundant water. Wash contaminated objects before reusing them.
- For information on hazardous substances included in the kit please refer to Material Safety Data Sheets (MSDS). The Material Safety Data Sheet for this product is available directly on the website of the manufacturer or upon request.
- Kit reagents must be regarded as hazardous waste and disposed of according to national regulations.

4. Storage and stability

Store the reagents at 2 - 8 °C until expiration date. Do not use components beyond the expiration date indicated on the kit labels. Do not mix various lots of any kit component within an individual assay.

5. Materials

5.1 Contents of the kit

<u>REF</u>	<u>Symbol</u>	<u>Reagent</u>	<u>Content</u>	<u>Colour Code</u>	
BA D-0035	MB 48	Master Block	1 x 48 wells		ready for use
BA E-0030	WASH-CONC 50x	Wash Buffer Concentrate	1 x 20 ml	light purple	concentrate
BA E-0055	SUBSTRATE	Substrate	1 x 12 ml	black	ready for use, containing a solution of (TMB)
BA E-0080	STOP-SOLN	Stop Solution	1 x 12 ml	light grey	ready for use
FC E-3601	C⇌0 PPM	Control 0 ppm	1 x 4 ml	white	ready for use
FC E-3602	C⇌3 PPM	Control 3 ppm	1 x 4 ml	light yellow	ready for use
FC E-3603	C⇌10 PPM	Control 10 ppm	1 x 4 ml	orange	ready for use
FC E-3604	C⇌20 PPM	Control 20 ppm	1 x 4 ml	light green	ready for use
FC E-3605	C⇌30 PPM	Control 30 ppm	1 x 4 ml	light purple	ready for use
FC E-3606	C⇌50 PPM	Control 50 ppm	1 x 4 ml	dark blue	ready for use
FC E-3607	C⇌100 PPM	Control 100 ppm	1 x 4 ml	light grey	ready for use
FC E-3608	C⇌300 PPM	Control 300 ppm	1 x 4 ml	black	ready for use
FC E-3611	ACYL-BUFF	Acylation Buffer	2 x 50 ml	white	ready for use
FC E-3612	ACYL-REAG	Acylation Reagent	1 x 3 ml	brown	ready for use, yellow coloured
FC E-3631	HIS	Histamine Microtiter Strips	1 x 48 wells		6 strips, 8 wells each, break apart, precoated
FC E-3640	HIS AB CONJ	Histamine Antiserum Conjugate	1 x 6 ml	red	ready for use, goat-anti Histamine IgG conjugated with peroxidase

5.2 Additional materials and equipment required but not provided with the kit

- Precision pipette (50 µl)
- Pipette tips (50 µl)
- Manual repetitive pipette (e.g. the Brand HandyStep® S)
- Precision Dispenser Tips (5 ml, 25 ml; e.g. the PLASTIBRAND® PD-Tips)
- Grinder (mill) or house hold blender
- Graduated plastic cylinder (250 ml)
- Water (deionized, distilled, or ultra-pure)
- Scale (capable of weighing 5 – 50 grams, precision 0.1 gram)
- Funnel and filter paper (or alternatively a centrifuge)
- Timer
- Waterproof marker
- Absorbent material (paper towel)
- Microplate Vibration Shaker (shaking amplitude 2 mm; approx. 600 rpm, (e.g. PSU-2T Minishaker *))
- ELISA reader capable of reading absorbance at 450 nm (required for semi-quantitative and quantitative determination)
- Washing device (plate washer or manually)

* Available upon request!

6. Test procedure

6.1 Preparation of reagents

Wash Buffer

Dilute the 20 ml Wash Buffer Concentrate with water (deionized, distilled, or ultra-pure) to a final volume of 1000 ml. Storage: up to 6 months at 2–8°C.

Acylation Reagent

The Acylation Reagent has a freezing point of 18.5 °C. To ensure that the Acylation Reagent is liquid when being used, it must be ensured that the Acylation Reagent has reached room temperature and forms a homogeneous, crystal-free solution before being used.

6.2 Sample preparation

The following protocols for the sample preparations are based on the **AOAC Official Method 937.07**

Sampling should be performed according to national regulation.

A. FRESH FISH • FROZEN FISH

- Keep (fresh) fish frozen prior to analysis.
- Thaw samples under refrigeration or in cold water. Do **not** thaw the samples in a heated water bath. Discard draining.
- Once thawed, store the samples refrigerated (2 - 8 °C) prior to testing.

WHOLE FISH:

Clean, scale and eviscerate fish. In case of small fish 6 in. (≤ 15 cm), use 5 – 10 fish. In case of large fish, from each of ≥ 3 fish cut 3 cross-sectional slices 1 in. (2.5 cm) thick, 1 slice from just back of pectoral fins, 1 slice halfway between first slice and vent, and 1 slice just back of vent. Remove bone. Blend combined samples until homogenous.

FISH FILET:

Use entire piece. Blend until homogenous.

B. CANNED FISH and OTHER CANNED MARINE PRODUCTS

Place entire content of the can (meat and liquid) in a blender and blend until homogenous.

C. CANNED MARINE PRODUCTS PACKED in OIL, SAUCE, BRINE or BROTH

Drain for 2 minutes on number 8 sieve or dab away the fluid with a paper towel. Place the meat in a blender and blend until homogenous.

D. FISHMEAL

Mix sample until homogenous.

6.3 Extraction

- | |
|--|
| <ul style="list-style-type: none">• Weigh 10 g of prepared fish sample / fish meal, add 240 ml water (deionized, distilled, or ultra-pure) and homogenize*) for 1-2 minutes in a grinder or blender.
*): Instead of homogenization fish meal samples are stirred for 10 minutes at room temperature. |
| <ul style="list-style-type: none">• Filter the homogenate through folded filter paper (alternatively an aliquot of the homogenate can be centrifuged for 5 minutes at maximum speed). <i>If a lipid layer forms remove it by suction!</i> |
| <ul style="list-style-type: none">• Use 50 μl of the sample extract for the acylation. |

6.4 Histamine ELISA

For the subsequent steps (Acylation and ELISA) allow all reagents and samples to reach room temperature.

A. SEMI-QUANTITATIVE DETERMINATION

For the semi-quantitative determination select the desired cut-off you need from the controls provided with the kit.

The kit controls have the following concentrations:

control 3, 10, 20, 30, 50, 100 or 300 ppm.



B. QUANTITATIVE DETERMINATION

For the quantitative determination use the following controls provided with the kit:


control 0, 3, 10, 30, 100 and 300 ppm.

These 6 controls are used to establish the standard curve (please refer to section 7.)

6.4.1 Acylation

1.	Pipette 50 µl of control(s) and sample extracts into the respective wells of the Master Block .
2.	Add 1.5 ml of Acylation Buffer in 1 (!) pipetting step to all wells. <i>The use of a repetitive pipette together with a new Precision Dispenser Tip (25 ml, please refer to chapter 5.), is recommended.</i>
3.	Add 50 µl of Acylation Reagent to all wells. (Colour change from yellow to pink!) Continue without any delay with step 4.!  <i>The use of a repetitive pipette together with a new Precision Dispenser Tip (2.5 ml, please refer to chapter 5.), is recommended.</i>
4.	Incubate 5 min at RT (20 - 25 °C) on a shaker (approx. 600 rpm).  Make sure that mixing is complete (slight pink colour).
5.	Take 50 µl for the ELISA

6.4.2 Histamine ELISA

1.	Pipette 50 µl of the acylated control(s) and samples into the wells of the Histamine Microtiter Strips .
2.	Pipette 100 µl of the Histamine Antiserum Conjugate into all wells. <i>The use of a repetitive pipette together with a new Precision Dispenser Tip (5 ml, please refer to chapter 5.), is recommended.</i>
3.	Incubate 10 min at RT (20 - 25 °C) on a shaker (approx. 600 rpm).
4.	Discard or aspirate the contents of the wells. Wash the plate 3 x by adding 300 µl of Wash Buffer , discarding the content and blotting dry each time by tapping the inverted plate on absorbent material.
5.	Pipette 100 µl of the Substrate into all wells. <i>The use of a repetitive pipette together with a new Precision Dispenser Tip (5 ml, please refer to chapter 5.), is recommended.</i>
6.	Incubate for 10 min at RT (20 - 25 °C) on a shaker (approx. 600 rpm).  Avoid exposure to direct sunlight!
7.	Add 100 µl of the Stop Solution to each well and shake the microtiter plate to ensure a homogeneous distribution of the solution. <i>The use of a repetitive pipette together with a new Precision Dispenser Tip (5 ml, please refer to chapter 5.), is recommended.</i>
8.	Read the absorbencies of the solutions in the wells within 10 minutes using a microplate reader set to 450 nm .

7. Calculation of results

A. Semi-quantitative results:

If the absorbance of the sample is **higher** than that of the selected **Cut-Off Control**, the sample has **passed**.


If the absorbance of the sample is **lower** than that of the selected **Cut-Off Control**, the sample has **failed**.

B. Quantitative results:

The absorbance readings of the 6 controls (0, 3, 10, 30, 100 and 300 ppm) are used to establish a standard curve.

Plot the absorbance readings of the controls (y-axis, linear) against the corresponding control concentrations (x-axis, log) using a concentration of 0.001 ppm for the 0-control (*this alignment is mandatory because of the logarithmic presentation of the data*). For the curve fitting a non-linear regression has to be applied.

The concentrations of the samples can be read **directly** from this standard curve.

 If a sample is off-curve it has to be diluted with water 1:10 and re-assayed. The result obtained has to be multiplied by the dilution factor of 10.

8. Warranty

This test kit was produced according to the latest developments in technology and subjected to stringent internal and external quality control checks. Any alteration of the test kit or the test procedure as well as the usage of reagents from different charges may have a negative influence on the test results and are therefore not covered by warranty. The manufacturer is not liable for damages occurring during transit.

9. Application lists for different kind of fish samples

All fish samples tested so far are suitable for the **HistaSure™ ELISA^{Fast Track}**. The lists below depict some major applications in different matrices.

9.1 Species validated through AOAC Certification

Fish Species	Presentation
Tuna	- canned chunk light - fresh/frozen yellow fin
Mahi Mahi	- fresh/frozen
Sardines	- canned in oil
Fishmeal	

9.2 Species approved through in-house testing











Fish Species	Presentation
Mackerel	- smoked
Anchovy	- fresh - brined - in sauce
Shad	- dry salted - fermented
Herring	- smoked
Salmon	- smoked
Bonito	- lakerda

10. Assay characteristics

AOAC performance tested method for fresh/frozen yellowfin tuna, canned tuna-chunk light in water, frozen mahi mahi, canned sardines in oil and fish meal.

Analytical Specificity (Cross Reactivity)	Substance			Cross Reactivity (%) Histamine	
	Histamine			100	
	L-Tryptophan			nd	
	Tryptamine			nd	
	3-Methylhistamine			0.44	
	L-Histidine			nd	
	L-Tyrosine			nd	
	L-Phenylalanine			nd	
	Tyramine			0.69	
	Cadaverine			0.40	
	Spermine			nd	
	Putrescine			nd	
Trimethylamine			nd		
nd = not detectable					
Accuracy and Precision					
Recovery				Intra CV	
Sample	Fortification range (ppm)	Mean recovery (%)	Recovery range (%)	Mean CV (%) (n=7)	Range CV (%)
Fresh/Frozen Tuna	5.25 – 218.6	91.8	85.2 – 99.6	7.58	4.13 – 10.8
Canned Tuna	5.49 – 267.3	99.8	94.7 – 106.5	8.74	3.17 – 13.9
Frozen Mahi Mahi	6.38 – 199.0	87.3	79.1 – 103.1	6.19	2.97 – 9.56
Canned Sardines	5.27 – 249.9	87.6	76.8 – 99.7	5.65	1.80 – 9.22
Fish meal	9.4 – 244.2	86.0	79.0 – 93.9	4.89	2.09 – 7.92
		Mean (ppm)	n	Inter CV %	
Lot to Lot		25.8	3 lots	4.16	
Detection limits					
LOD (Limit of Detection) 0.44 ppm			LOQ (Limit of Quantification) 1.31 ppm		
Method Comparison					
LDN HistaSure Elisa vs AOAC 977.13 fluorometric method: Fresh/Frozen Tuna, Canned Tuna, Frozen Mahi Mahi and Canned Sardines					

Symbols:

	Contains sufficient for <n> tests		Manufacturer		Storage temperature
	Catalogue number		Batch code		Expiry date
	Caution		Content		Consult instructions for use
	For research use only!				