

Instructions for use
N1-Acetyl-Spermidine ELISA

REF

IS I-2400R



RUO

N1-Acetyl-Spermidine ELISA

1. Intended use and principle of the test

Enzyme Immunoassay for the quantitative determination of N1-Acetyl-Spermidine (N1-actSp) in plasma samples.

After precipitation and derivatization N1-Acetyl-Spermidine is quantitatively determined by ELISA.

The competitive ELISA uses the microtiter plate format. The antigen is bound to the solid phase of the microtiter plate. The processed standards, controls and samples and the solid phase bound analyte compete for a fixed number of antiserum binding sites. When the system is in equilibrium, free antigen and free antigen-antiserum complexes are removed by washing. The antibody bound to the solid phase is detected by an anti-rabbit IgG-peroxidase conjugate using TMB as a substrate. The reaction is monitored at 450 nm.

Quantification of unknown samples is achieved by comparing their absorbance with a reference curve prepared with known standards.

2. Procedural cautions, guidelines, warnings and limitations

2.1 Procedural cautions, guidelines and warnings

- (1) This kit is intended for professional use only. Users should have a thorough understanding of this protocol for the successful use of this kit. Only the test instruction provided with the kit is valid and has to be used to run the assay. Reliable performance will only be attained by strict and careful adherence to the instructions provided.
- (2) This assay was validated for certain types of samples as indicated in *Intended Use* (please refer to Chapter 1). Any off-label use of this kit is in the responsibility of the user and the manufacturer cannot be held liable.
- (3) The principles of Good Laboratory Practice (GLP) have to be followed.
- (4) In order to reduce exposure to potentially harmful substances, wear lab coats, disposable protective gloves and protective glasses where necessary.
- (5) All kit reagents and specimens should be brought to room temperature and mixed gently but thoroughly before use. Avoid repeated freezing and thawing of reagents and specimens.
- (6) For dilution or reconstitution purposes, use deionized, distilled or ultra-pure water.
- (7) The microplate contains snap-off strips. Unused wells must be stored at 2 °C to 8 °C in the sealed foil pouch with desiccant and used in the frame provided.
- (8) Duplicate determination of sample is highly recommended to be able to identify potential pipetting errors.
- (9) Once the test has been started, all steps should be completed without interruption. Make sure that the required reagents, materials and devices are prepared ready at the appropriate time.
- (10) Incubation times do influence the results. All wells should be handled in the same order and time intervals.
- (11) To avoid cross-contamination of reagents, use new disposable pipette tips for dispensing each reagent, sample, standard and control.
- (12) A standard curve must be established for each run.
- (13) The controls should be included in each run and fall within established confidence limits. The confidence limits are listed in the QC-Report.
- (14) Do not mix kit components with different lot numbers within a test and do not use reagents beyond expiry date as shown on the kit labels.
- (15) Avoid contact with Stop Solution containing 0.25 M H₂SO₄. It may cause skin irritation and burns. In case of contact with eyes or skin, rinse off immediately with water.
- (16) TMB substrate has an irritant effect on skin and mucosa. In case of possible contact, wash eyes with an abundant volume of water and skin with soap and abundant water. Wash contaminated objects before reusing them.
- (17) For information on hazardous substances included in the kit please refer to Safety Data Sheet (SDS). The Safety Data Sheet for this product is made available directly on the website of the manufacturer or upon request.
- (18) The expected reference values reported in this test instruction are only indicative. It is recommended that each laboratory establishes its own reference intervals.
- (19) The results obtained with this test kit should not be taken as the sole reason for any therapeutic consequence but have to be correlated to other diagnostic tests and clinical observations.
- (20) Kit reagents must be regarded as hazardous waste and disposed of according to national regulations.

2.2 Limitations

Any inappropriate handling of samples or modification of this test might influence the results.


Samples containing precipitates or fibrin strands or which are haemolytic or lipemic might cause inaccurate results.

3. Storage and stability

Store the unopened reagents at 2 - 8 °C until expiration date. Do not use components beyond the expiry date indicated on the kit labels. Once opened the reagents are stable for 1 month when stored at 2 - 8 °C. Once the resealable pouch has been opened, care should be taken to close it tightly with desiccant again.

4. Materials

4.1 Contents of the kit

- BA D-0090** **FOILS** **Adhesive Foil** - Ready to use
Contents: Adhesive Foils in a resealable pouch
Volume: 1 x 4 foils
- IS I-0324** **REAC-PLATE** **Reaction Plate** - Ready to use
Contents: 1 x 96 well plate, empty in a resealable pouch
- BA E-0030** **WASH-CONC 50x** **Wash Buffer Concentrate** - Concentrated 50x
Contents: Buffer with a non-ionic detergent and physiological pH
Volume: 1 x 20 mL/vial, light purple cap
- BA E-0040** **CONJUGATE** **Enzyme Conjugate** - Ready to use
Contents: Goat anti-rabbit immunoglobulins conjugated with peroxidase
Volume: 1 x 12 mL/vial, red cap
- BA E-0055** **SUBSTRATE** **Substrate** - Ready to use
Contents: Chromogenic substrate containing tetramethylbenzidine, substrate buffer and hydrogen peroxide
Volume: 1 x 12 mL/black vial, black cap
- BA E-0080** **STOP-SOLN** **Stop Solution** - Ready to use
Contents: 0.25 M sulfuric acid
Volume: 1 x 12 mL/vial, light grey cap
Hazards identification: 
H290 May be corrosive to metals.
- IS I-2431** **IT1 N1-actSp** **N1-Acetyl-Spermidine Microtiter Strips** - Ready to use
Contents: 1 x 96 well (12x8) antigen precoated microwell plate in a resealable pouch with desiccant, grey bag
- IS I-2410** **AS N1-actSp** **N1-Acetyl-Spermidine Antiserum** - Ready to use
Contents: Rabbit anti-N1-Acetyl-Spermidine antibody, green coloured
Volume: 1 x 6 mL/vial, yellow cap
- BA E-2413** **ASSAY-BUFF** **Assay Buffer** - Ready to use
Contents: Buffer with alkaline pH
Volume: 1 x 20 mL/vial, yellow cap
- BA E-2428** **EQUA-REAG** **Equalizing Reagent** - Lyophilized
Contents: Lyophilized protein
Volume: 1 vial, brown cap
- BA E-2446** **D-REAGENT** **D-Reagent** - Ready to use
Contents: Crosslinking agent in dimethylsulfoxide
Volume: 1 x 3 mL/vial, white cap

BA E-2458 **Q-BUFFER** **Q-Buffer** - Ready to use

Contents: TRIS buffer
 Volume: 1 x 20 ml/vial, white cap

BA E-2788 **PBS** **PBS** - Ready to use

Contents: Phosphate Buffered Saline
 Volume: 1 x 20 ml/vial, orange cap

BA E-2721 **PREC-REAG** **Precipitating Reagent** - Ready to use

Contents: Acidic reagent for precipitation of plasma/serum proteins, red coloured
 Volume: 1 x 4 mL/vial, white cap

Standards and Controls - Ready to use

Cat. no.	Component	Colour/Cap	Concentration ng/mL	Concentration nmol/L	Volume/ Vial
IS I-2401	STANDARD A	white	0	0	4 mL
IS I-2402	STANDARD B	light yellow	2,08	8	4 mL
IS I-2403	STANDARD C	orange	5,20	20	4 mL
IS I-2404	STANDARD D	dark blue	13,01	50	4 mL
IS I-2405	STANDARD E	light grey	32,52	125	4 mL
IS I-2406	STANDARD F	black	81,31	312,5	4 mL
IS I-2451	CONTROL 1	light green	Refer to QC-Report for expected value and acceptable range!		4 mL
IS I-2452	CONTROL 2	dark red			4 mL

Conversion: N1-Acetyl-Spermidine (ng/mL) x 3,843 = N1-Acetyl-Spermidine (nmol/L)

Contents: Neutral buffer with non-mercury stabilizer, spiked with defined quantity of N1-Acetyl-Spermidine

4.2 Additional materials and equipment required but not provided in the kit

- Calibrated precision pipettes to dispense volumes between 10 – 300 µL; 12.5 mL
- Polystyrene or polypropylene tubes (0.5 mL) and suitable rack
- Microtiter plate washing device (manual, semi-automated or automated)
- ELISA reader capable of reading absorbance at 450 nm and if possible 620 - 650 nm
- Microtiter plate shaker (shaking amplitude 3 mm; approx. 600 rpm)
- Absorbent material (paper towel)
- Water (deionized, distilled or ultra-pure)
- Vortex mixer

5. Sample collection and storage**EDTA Plasma**

Whole blood should be collected into centrifuge tubes containing EDTA as anti-coagulant (Monovette™ or Vacuette™ for plasma) and centrifuged according to manufacturer's instructions at room temperature immediately after collection.

Haemolytic and especially lipemic samples should not be used for the assay.

Storage: up to 48 hours at 2 - 8 °C, for longer period (up to 6 month) at -20 °C.

Repeated freezing and thawing should be avoided.

6. Test procedure

Allow all reagents and samples to reach room temperature and mix thoroughly by gentle inversion before use. Duplicate determinations are recommended. To do that, perform duplicates before derivatization (1 precipitation tube = 2 derivatization wells).

It is recommended to number the strips of the microwell plate before usage to avoid any mix-up.

The binding of the antisera and of the enzyme conjugate and the activity of the enzyme are temperature dependent, and the absorbance values may vary if a thermostat is not used. The higher the temperature, the higher the extinction values will be. Corresponding variations also apply to the incubation times. The optimal temperature during the Enzyme Immunoassay is between 20 – 25 °C.

6.1 Preparation of reagents

Wash Buffer

Dilute the 20 mL Wash Buffer Concentrate with water (deionized, distilled or ultra-pure) to a final volume of 1000 mL.

Storage: 1 month at 2 – 8 °C

Equalizing Reagent

Reconstitute the Equalizing Reagent with **12.5 mL** of **Assay Buffer**.

Reconstituted Equalizing Reagent which is not used immediately has to be stored in aliquots for max 1 month at -20 °C and may be thawed only once.


D-Reagent

The D-Reagent has a freezing point of 18.5 °C. It must be ensured that the D-Reagent has reached room temperature and forms a homogeneous, crystal-free solution.


N1-Acetyl-Spermidine Microtiter Strips

In rare cases residues of the blocking and stabilizing reagent can be seen in the wells as small, white dots or lines. These residues do not influence the quality of the product.


6.2 Precipitation

1.	Pipette 50 µL of standards, controls and samples into the respective tubes.
2.	Add 50 µL of PBS to the standards, controls and samples.
3.	Add 10 µL of Precipitating Reagent to all tubes.
4.	Mix the tubes thoroughly (vortex) and centrifuge for 15 minutes at 3000 x g .
	Take 25 µL of the clear supernatant for the derivatization .

6.3 Derivatization

1.	Pipette 25 µL of the precipitated standards, controls and samples into the appropriate wells of the Reaction Plate .
2.	Add 50 µL of the Equalizing Reagent into all wells (2 min at RT under stirring at 700 rpm)
3.	Add 10 µL of the D-Reagent into all wells (immerse the tips into the reaction medium).
4.	Cover plate with Adhesive Foil and incubate for 2 h at RT (20 – 25 °C) on a shaker (approx. 500 rpm).
5.	Add 100 µL of the Q-Buffer into all wells.
6.	Incubate for 10 min at RT (20 – 25 °C) on a shaker (approx. 500 rpm).
	Use 100 µL for the ELISA!

6.4 N1-Acetyl-Spermidine ELISA

1.	Pipette 100 µL of the prepared standards, controls and samples into the appropriate wells of the N1-Acetyl-Spermidine Microtiter Strips . Be careful not to pipette bubbles!
2.	Add 50 µL of the N1-Acetyl-Spermidine Antiserum into all wells and mix shortly.
3.	Cover plate with Adhesive Foil and incubate for 15 - 20 h (overnight) at 2 – 8 °C .
4.	Remove the foil. Discard or aspirate the content of the wells. Wash the plate 4 x by adding 300 µL of Wash Buffer , discarding the content and blotting dry each time by tapping the inverted plate on absorbent material.
5.	Pipette 100 µL of the Enzyme Conjugate into all wells.
6.	Incubate for 30 min at RT (20 – 25 °C) on a shaker (approx. 500 rpm).
7.	Discard or aspirate the content of the wells. Wash the plate 4 x by adding 300 µL of Wash Buffer , discarding the content and blotting dry each time by tapping the inverted plate on absorbent material.
8.	Pipette 100 µL of the Substrate into all wells and incubate for 15 - 25 min at RT (20 – 25 °C) on a shaker (approx. 500 rpm). Avoid exposure to direct sunlight!
	

9. Add **100 µL** of the **Stop Solution** to each well and shake the microtiter plate to ensure a homogeneous distribution of the solution.
10. **Read the absorbance** of the solution in the wells within 10 minutes, using a microplate reader set to **450 nm** (if available a reference wavelength between 620 nm and 650 nm is recommended).

7. Calculation of results

Measuring range	N1-Acetyl-Spermidine
	8 – 312,5 nM

The calibration curve is obtained by plotting the absorbance readings (calculate the mean absorbance) of the standards (linear, y-axis) against the corresponding standard concentrations (logarithmic, x-axis). Use non-linear regression for curve fitting (e.g. spline, 4- parameter, akima).

⚠ *This assay is a competitive assay. This means: the OD-values are decreasing with increasing concentrations of the analyte. OD-values found below the standard curve correspond to high concentrations of the analyte in the sample.*

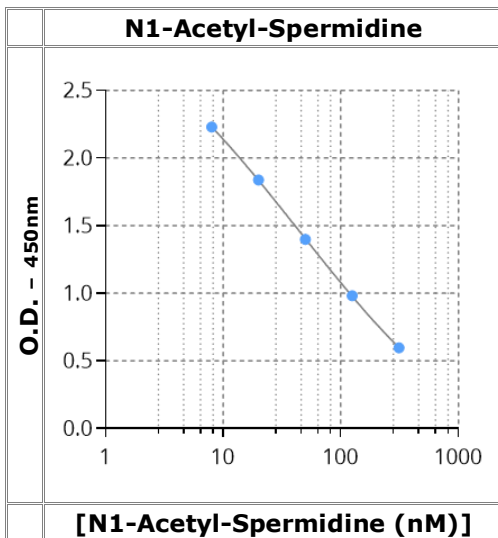
The concentrations of the samples and controls can be read directly from the standard curve.

7.1 Quality control

The confidence limits of the kit controls are indicated on the QC-Report.

7.2 Typical standard curve

⚠ *Example, do not use for calculation!*



8. Assay characteristics

Analytical Sensitivity (Limit of Detection)	N1-Acetyl-Spermidine
	3,5 nM


Analytical Specificity (Cross Reactivity)	Substance	Cross Reactivity (%)
	N1-Acetyl-Spermidine	100%
	N8-Acetyl-Spermidine	1,7%
	Spermidine	<0,1%
	Spermine	<0,1%
	Ornithine	<0,1%
	Putrescine	<0,1%

Precision			
Intra-Assay			
Plasma Sample	Mean (nM)	SD (nM)	CV (%)
1 (n = 8)	31	4	12
2 (n = 8)	76	9	9
3 (n = 8)	139	15	10







Linearity	Plasma Sample (Serial dilution up to 1:32)	Range Linearity (%)	Mean Linearity (%)
	1	90 - 109	102
	2	92 - 124	108
	3	85 - 124	96

Recovery	Plasma Sample	Range Recovery (%)	Mean Recovery (%)
	1	93 - 106	100
	2	94 - 102	98
	3	99 - 104	102

Method Comparison: ELISA vs LC/MS	Plasma samples	$[N1\text{-Acetyl-Spermidine}]_{ELISA} = 0,6554 * [N1\text{-Acetyl-Spermidine}]_{LC-MS} + 10,21$	$R^2 = 0,9753$ N = 40
--	----------------	--	--------------------------

 For updated literature or any other information please contact your local supplier.

Symbols:

 $+2$ $^{+8}$ $^{\circ}C$	Storage temperature		Manufacturer		Contains sufficient for <n> tests
	Expiry date	LOT	Batch code	RUO	For research use only!
	Consult instructions for use	CONT	Content		
	Caution	REF	Catalogue number		